

gndDb, a Database of Partial *gnd* Sequences To Assist with Analysis of *Escherichia coli* Communities Using High-Throughput Sequencing

Microbiology

Resource Announcements

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ABSTRACT The use of culture methods to detect *Escherichia coli* diversity does not provide sufficient resolution to identify strains present at low levels. Here, we target the hypervariable *gnd* gene and describe a database containing 534 distinct partial *gnd* sequences and associated O groups for use with culture-independent *E. coli* community analysis.

Current culture-dependent studies to investigate *Escherichia coli* diversity in fecal and environmental samples often fail to identify strains that are present in low numbers. Our previous work using sequencing of metabarcoded amplicons has targeted the hypervariable *gnd* gene to provide a comprehensive analysis of *E. coli* community structure from complex samples such as feces (1). The *gnd* gene encodes 6-phosphogluconate dehydrogenase, the third enzyme in the pentose phosphate pathway, and is found in most *Enterobacteriaceae* (2–4). Usually, *gnd* is found adjacent to the highly recombinatorial O-antigen biosynthesis gene cluster (O-AGC) (5, 6), a region of the *E. coli* chromosome prone to horizontal gene transfer and recombination (7), which influences the O-group structure and cell surface antigenicity as the outermost component of the lipopolysaccharide (LPS) moiety. Although having no role in O-antigen biosynthesis, *gnd* has been described as a passive hitchhiker of recombination events influencing LPS antigenic changes (4).

By targeting *gnd* polymorphisms and adopting culture-independent methods, our previous work provided an indication of intestinal *E. coli* diversity and in parallel developed a *gnd* database for cross-referencing purposes using O-AGC DNA sequence data from distinct O groups (1). However, the increasing availability and analysis of whole-genome sequencing (WGS) data from *E. coli* (and *Shigella*) isolates have provided new insights as to the range of different *E. coli* O groups according to incongruent *wzx* and *wzy* (provisional OXY designations) or *wzm* and *wzt* (provisional OMT designations) gene sequences and the identification of six novel O groups (8). By isolating and examining the *gnd* gene from *E. coli* and *Shigella* draft genome assemblies described in recent studies (9, 10) and in other studies where novel O-AGCs have been submitted to GenBank (5, 6, 8), we have identified novel *gnd* alleles that have been included in a database resource that may be used for analysis of *E. coli* communities using amplicon sequencing. Analysis of WGS data from *E. coli* (and *Shigella*) with novel O groups has

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Received 25 April 2019 Accepted 18 July 2019 Published 15 August 2019 provided evidence for the identification of 534 distinct *gnd* sequence types (gSTs), each 284 bp in length, forming the core of this database.

The full-length *gnd* gene of 1,407 bp precludes its use in its entirety as a tool for culture-independent amplicon sequencing studies using the Illumina platform; therefore, the 284-bp region spanning nucleotide positions 443 to 727 is targeted for use in *E. coli* community analysis studies. The hypervariable nature of *gnd* also restricts suitable PCR primer sites for the generation of amplicons of a suitable size for routine high-throughput sequencing using the Illumina platform. This *gnd* database may also provide some assistance in the identification of novel O groups and offer a resource for *E. coli* subtyping using conventional dideoxy Sanger sequencing methods as a primary screen for subsequent WGS analysis (11).

Data availability. The DNA sequences of the 534 distinct gSTs are available in FASTA format from GitHub (https://github.com/mEpiLab/gnd) and are accompanied by a spreadsheet which provides matching O groups for each gST and a representative accession number of matching draft genome assemblies or submitted nucleotide sequences. The database and list of matching O groups will be curated and updated as further WGS data are made available.

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